

Running Head: Heuristic climate bio-regulation modelling

Dimethylsulphide, clouds and phytoplankton: insights
from a simple plankton ecosystem feedback model.

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18 **ABSTRACT**

19 The hypothesis that marine plankton ecosystems may effectively regulate climate by
20 the production of dimethylsulphide (DMS) has attracted substantial research effort
21 over recent years. This hypothesis suggests that DMS produced by marine ecosystems
22 can affect cloud properties and hence the averaged irradiance experienced by the
23 phytoplankton that produce DMS's precursor dimethylsulphoniopropionate (DMSP).
24 This paper describes the use of a simple model to examine the effects of such a
25 biogenic feedback on the ecosystem that initiates it. We compare the responses to
26 perturbation of a simple marine nitrogen – phytoplankton – zooplankton (NPZ)
27 ecosystem model with and without biogenic feedback. Our analysis of this heuristic
28 model reveals that the addition of the feedback can increase the model's resilience to
29 perturbation and hence stabilise the model ecosystem. This result suggests the
30 hypothesis that DMS may play a role in stabilising marine plankton ecosystem
31 dynamics through its effect on the atmosphere.

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33 **INDEX TERMS:** 0315, 0439, 0444, 0466

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35 **KEYWORDS:** DMS, climate change, CLAW, biofeedback, NPZ model.

36

37 **INTRODUCTION**

38 The prospect of human-induced climate change has stimulated research into several
39 biological processes that might affect climate. One such process that has attracted a
40 substantial research effort is the so-called CLAW hypothesis [*Charlson, et al.*, 1987].

This hypothesis suggests that marine plankton ecosystems may effectively regulate climate by a feedback associated with the production of dimethylsulphide (DMS). Charlson et al. [1987] observed that some of the DMS produced by marine ecosystems is transferred from the ocean to the atmosphere where it is the major source of cloud condensing nuclei (CCN) over the remote oceans. The aerosols resulting from biogenic DMS emissions can have a direct effect on the solar radiative forcing experienced by the Earth through scattering, absorption and reflection and can also lead to increased cloud formation; the CLAW hypothesis proposes that these mechanisms could regulate climate. Charlson et al [1987] argued that an increase in global temperature would lead to increased biogenic DMS emissions from the ocean and result in an increase in scattering, cloud cover and cloud albedo that would increase the proportion of the incoming solar radiation reflected back into space (thus changing the global albedo), and thereby cooling the planet.

DMS is an ecosystem product. Many species of marine phytoplankton synthesise dimethylsulphoniopropionate (DMSP), the precursor to DMS. However, most DMSP that is converted to DMS is done so by ecosystem processes that occur outside the phytoplankton cell [*Simo*, 2001]. In addition to the climatic role postulated for DMS by the CLAW hypothesis, DMSP is a compatible solute that can protect cells from the osmotic effects of seawater and the effects of freezing, may deter predation by zooplankton, may act as an antioxidant, and has also been proposed to assist in the long range dispersal of marine phytoplankton [*Hamilton and Lenton*, 1998; *Kiene, et al.*, 2000; *Sunda, et al.*, 2002; *Wolfe, et al.*, 1997].

The objective of this paper is to examine the implications of the climate regulation process proposed by Charlson et al. [1987] for the dynamics of the ecosystems that produce it. To do this we develop a simple plankton model that incorporates the DMS feedback mechanism and compare its dynamics to the same ecosystem model without the feedback. The plankton model is composed of nutrient (N), phytoplankton (P) and zooplankton (Z), and is one of a class of NPZ models that have proved powerful heuristic tools in biological oceanography [Franks, 2002]. The feedback processes quantified by Charlson et al. [1987] (a changed light environment arising from a modified albedo due to DMS production, where the changed light environment then influences the phytoplankton growth rate) were incorporated into the NPZ model to build a biogenic feedback model that was not intrinsically constrained to any specific oceanic environment. The structure of this model is shown in Figure 1. Our whole-of-system modelling approach is similar to that of Lawrence [1993], who used a similar model to assess the impact of the feedback on climate. Our approach and intention is substantially different: we close the feedback cycle on the ecosystem, and examine the implications of the feedback for the ecosystems that generate it.

Comparisons between simulations of the NPZ model with and without feedback were undertaken to elucidate the influence of the feedbacks for the ecosystem. The simulations revealed that the presence of the feedback generally enhanced the stability of the ecosystem by making it more resilient to perturbation. Resilience is a form of stability that may be analytically evaluated for many simple ecosystem models. The resilience of model ecosystems is defined as the negative of the dominant eigenvalue of the linearised system about the steady state [DeAngelis, 1980], and is inversely

related to the time the model ecosystem will take to return to its equilibrium state after a perturbation.

A sensitivity analysis of the model with instantaneous feedback (the reason we examine instantaneous feedback is discussed) indicated that such feedback always stabilised the system, that is no parameter values or combination of parameter values used in the sensitivity analysis caused the system to become less stable than the equivalent system without the feedback. We note that the feedback system is more sensitive to the magnitude and direction of zooplankton perturbations than to phytoplankton perturbations. We also note the sensitive dependence of the feedback effect on time lags in the feedback process, and the influence of the biometric rate parameters on these effects. These parameters are also important determinants of the resilience of the ecosystem model

THE ECOSYSTEM MODEL

We first describe the basic *NPZ-DMS* model without feedback, summarised in equations (1) - (5), and the methods used in this analysis. We then develop the feedback model (equation (34) and equations (35) - (37)).

The *NPZ-DMS* model

A simple model of a planktonic ecosystem that produces DMS is used as the starting point for this analysis. The model is a simplification of the GMSK model proposed by Gabric et al. [1993], in which a DMS - DMSP model was coupled to the plankton ecosystem model of Moloney et al. [1986]. This model is shown in Figure 1, where it comprises all the model components in the ocean. The ecosystem model has been simplified to a nutrient – phytoplankton - zooplankton (*NPZ*) model, a valuable tool in biological oceanography [Franks, 2002], for this analysis and coupled to the DMS equations of Gabric et al. [1993]. A schematic of the resulting simple *NPZ-DMS* model comprises the model components shown as solid lines in the ocean in Figure 1.

The *NPZ-DMS* model is defined by the following equations:

$$\frac{dN}{dt} = k_5 Z + k_3 k_4 PZ - k_1 \left(\frac{N}{N + k_2} \right) P, \quad (1)$$

$$\frac{dP}{dt} = k_1 \left(\frac{N}{N + k_2} \right) P - k_3 PZ, \quad (2)$$

$$\frac{dZ}{dt} = k_3 (1 - k_4) PZ - k_5 Z, \quad (3)$$

$$\frac{dDMSP}{dt} = m_1 \gamma P + m_2 \gamma Z - m_3 DMSP - m_4 DMSP, \quad (4)$$

$$\frac{dDMS}{dt} = \gamma m_5 P + m_3 DMSP - m_6 DMS - m_7 DMS. \quad (5)$$

127

128 Equations (1) - (3) represent the ecosystem component of the model. These equations
 129 are written in a currency of nitrogen (where N , P and Z represent the concentrations of
 130 dissolved inorganic nutrient, phytoplankton and zooplankton respectively in units of
 131 atomic nitrogen). These equations conserve mass so that $N + P + Z = N_T$ always,
 132 where N_T is the total nutrient in the system. Equations (4) and (5) represent the DMS
 133 production generated by the ecosystem and are written in a currency of sulphur
 134 (where $DMSP$ and DMS represent the concentrations of dimethylsulphoniopropionate
 135 and dimethylsulphide respectively in units of atomic sulphur) and these equations do
 136 not conserve mass. The k and m parameters represent the attributes of biota in the
 137 ecosystem model and the rates of chemical reactions and physical processes in the
 138 DMS model. The parameter γ in equations (4) and (5) represents the sulphur :
 139 nitrogen ratio of marine plankton. The parameter set used by Gabric et al. [1999],
 140 derived from measured values reported in the literature, was selected as a starting
 141 point for this analysis (Table 1, G99). The equations derived by Gabric et al [1993]
 142 were used as the basis for developing a feedback process to incorporate in the *NPZ*
 143 model to create the biofeedback model. This is described below in The Feedback
 144 Model.

145

146 ***NPZ* model resilience**

147 The nitrogen-based components of the *NPZ-DMS* model may be isolated to form an
 148 ecosystem (*NPZ*) model. This model has four critical (equilibrium) points defined by

149 $\frac{dN}{dt} = \frac{dP}{dt} = \frac{dZ}{dt} = 0$ of which one is ecologically infeasible and two are unstable and

not relevant to this analysis (see Cropp and Norbury [2007] for details). The fourth point is an asymptotically stable node located at:

$$P^* = \frac{k_5}{k_3(1-k_4)}, \quad (6)$$

$$Z^* = \frac{k_1}{k_3} \left(\frac{N^*}{N^* + k_2} \right), \quad (7)$$

$$N^* = \frac{1}{2} \left[- \left(k_2 + P^* + \frac{k_1}{k_3} - N_T \right) \pm \sqrt{\left(k_2 + P^* + \frac{k_1}{k_3} - N_T \right)^2 - 4k_2(P^* - N_T)} \right]. \quad (8)$$

The eigenvalues of the linearised NPZ model about this point are:

$$\lambda = -\frac{k_1 P^*}{2} \left[\frac{k_2}{(N^* + k_2)^2} \right] \left\{ 1 \pm \sqrt{1 - (N^* + k_2) \left[1 + \frac{(N^* + k_2)^2}{k_1 k_2 P^*} \right] \left(\frac{4k_4 N^*}{k_2 P^*} \right)} \right\}. \quad (9)$$

For the parameter values used in this study (and for most reasonable values) this point is an asymptotically stable spiral node, indicating that when the model is perturbed from this steady state it will oscillate in “boom-and-bust” cycles of decreasing amplitude until it regains equilibrium. The amplitude and rate of decay of these cycles

is determined by the magnitude of the real part of the eigenvalue of this point, the basis of DeAngelis' [1980] resilience measure:

$$Res = \frac{k_1}{2} \left(\frac{k_2}{(N^* + k_2)^2} \right) \left(\frac{k_5}{k_3(1 - k_4)} \right). \quad (10)$$

This resilience metric is inversely proportional to the time required for the model to return to equilibrium after a perturbation and we will use this metric to describe the stability of the model.

High resilience means that a system will rapidly return to its steady state after a perturbation. Johnson [1990] studied isolated Arctic lake ecosystems and proposed the hypothesis that ecosystems adapt to acquire attributes that endow the system with high resilience. Cropp and Gabric [2002] simulated the adaptation of a simple plankton ecosystem model under thermodynamic constraints and modified Johnson's hypothesis to propose that ecosystems adapted to states of maximum resilience within genetic and environmental constraints. Laws [2003] reported that the concept of maximum resiliency was a useful heuristic for fitting ecosystem models to observed plankton data in many regions of the global oceans.

THE FEEDBACK MODEL

The feedback model is based on the simplified GMSK model and is comprised of the *NPZ* model described above coupled to a model of the sulphur and atmospheric processes. The feedback model is also shown in Figure 1, and comprises all the model components drawn with solid lines in both the ocean and the atmosphere.

A *DMS* sub-model may be extracted from the sulphur-based components of the *NPZ-DMS* model (equations (4) - (5)) for analysis. A steady state of the *DMS* sub-model may be obtained where the aqueous *DMS* concentration is defined in terms of P , Z and some parameters:

$$DMS^* = \gamma \left(\frac{P(m_1 m_3 + m_5 m_3 + m_5 m_4) + m_2 m_3 Z}{(m_3 + m_4)(m_6 + m_7)} \right). \quad (11)$$

This steady state is always an asymptotically stable node, and for measured parameter values the eigenvalues of this point are large and negative, indicating that the steady state is a strong attractor and the *DMS* model is highly resilient. This means that the system will return to its steady state very rapidly after any perturbation, and that the steady state values are a therefore a good description of the system. Analytic expressions describing $P(t)$ and $Z(t)$ are not derivable, but it is clear that the *DMS* model is essentially slaved to the *NPZ* model, and the state of the ecosystem model will determine the state of the *DMS* model. These properties of the *NPZ-DMS* model

allow the effects of the system's feedback on its own environment to be included in a simple ecosystem biofeedback model. The derivation of this model is now described.

DMS sea-air transfer

The flux of DMS from the ocean to the atmosphere (DMS_{flux}) may be modeled as a linear function of the aqueous concentration of DMS (DMS_{aq}) and the DMS piston velocity (k_{tr}) [Liss and Merlivat, 1986]:

$$DMS_{flux} = k_{tr} DMS_{aq} . \quad (12)$$

Although this flux occurs only at the ocean surface, when applied in a zero-dimensional (i.e. depth-averaged over the mixed layer) model as in this analysis, the loss of DMS_{aq} is also averaged over the depth of the mixed layer (MLD):

$$DMS_{flux} = \left(\frac{k_{tr}}{MLD} \right) DMS_{aq} . \quad (13)$$

The DMS piston velocity parameterises the rate at which DMS is transferred from the ocean to the atmosphere, and the empirically derived estimate of Nightingale et al. [2000] was used in this analysis:

225

$$k_{tr} = 0.005(5.88u_{10}^2 + 1.49u_{10})Sc^{-1/2}, \quad (14)$$

227

228 where k_{tr} (day^{-1}) is for a notional 48 metre deep mixed layer, u_{10} is the 10 metre wind
 229 speed (ms^{-1}) and Sc is the dimensionless Schmidt number. The Schmidt number is
 230 temperature dependent, and the experimentally derived relationship of Saltzman et al.
 231 [1993] was used:

232

$$Sc = 2764 - 147.12 SST + 3.726 SST^2 - 0.038 SST^3, \quad (15)$$

234

235 where SST is the sea surface temperature ($^{\circ}\text{C}$). The sea surface temperature has little
 236 influence on the piston velocity, which is mostly controlled by the wind speed. A
 237 global average wind speed of 8.2 ms^{-1} (derived from two years of SeaWinds 12 hourly
 238 measurements) and a global average sea surface temperature of 16.7°C (derived from
 239 15 years of Advanced Very High Resolution Radiometer (AVHRR) data) were used
 240 for this analysis, giving a representative DMS piston velocity of $k_{tr} = 2.88 \text{ md}^{-1}$. This
 241 is consistent with the estimates of Simo and Dachs [2002] who used a non-linear
 242 correction to zonal monthly climatological wind speeds and obtained global estimates
 243 of piston velocity between 1 and 4 md^{-1} .

244

A relationship between P , Z and DMS flux from the ocean to the atmosphere can be derived by substituting equation (11) into equation (12):

$$DMS_{flux} = k_{tr}\gamma \left[\frac{(m_1m_3 + m_3m_5 + m_4m_5)}{(m_3 + m_4)(m_6 + m_7)} P + \frac{m_2m_3}{(m_3 + m_4)(m_6 + m_7)} Z \right], \quad (16)$$

$$= k_6P + k_7Z$$

where $k_6 = \frac{\gamma k_{tr}(m_1m_3 + m_3m_5 + m_4m_5)}{(m_3 + m_4)(m_6 + m_7)}$, and $k_7 = \frac{\gamma k_{tr}m_2m_3}{(m_3 + m_4)(m_6 + m_7)}$ where k_6 and k_7

have units of $\text{mgS m mgN}^{-1} \text{ day}^{-1}$ and DMS_{flux} has units of $\text{mgS m}^{-2} \text{ day}^{-1}$.

DMS flux and cloud condensation nuclei

Pandis et al. [1994] estimated a long-term average cloud condensation nuclei (CCN) - DMS flux relationship over the remote ocean from the steady state of a model of the principal gas-, aerosol- and aqueous-phase processes in the marine atmospheric boundary layer. They predicted that when DMS emission flux is smaller than $1.3 \mu\text{moles m}^{-2} \text{ d}^{-1}$ the CCN concentration is essentially constant at $20 \text{ particles cm}^{-3}$. In a second DMS flux regime, extending between 1.3 and $2.3 \mu\text{moles m}^{-2} \text{ d}^{-1}$ only a few particles become CCN each day. A third region corresponds to DMS emission fluxes larger than $2.3 \mu\text{moles m}^{-2} \text{ d}^{-1}$. The CCN concentration in this regime is a linear function of the DMS flux:

$$CCN = 22.7 DMS_{flux} - 15.0, \quad (17)$$

265

266 where DMS emission flux is in $\mu\text{moles m}^{-2} \text{ d}^{-1}$ and CCN is in particles cm^{-3} . Simo and
 267 Dachs [2002] estimated a conservative global mean annual flux of DMS from the
 268 ocean to the atmosphere of $6.8 \mu\text{moles m}^{-2} \text{ d}^{-1}$ ($\sigma = 2.49 \mu\text{moles m}^{-2} \text{ d}^{-1}$) suggesting
 269 that equation (17) is appropriate for a globally representative model. Converting this
 270 relationship to represent DMS in $\text{mg S m}^{-2} \text{ d}^{-1}$ using $1 \mu\text{mol S m}^{-2} \text{ d}^{-1} = 0.032 \text{ mgS m}^{-2}$
 271 d^{-1} gives approximately:

272

$$CCN = 710 DMS_{flux} - 15. \quad (18)$$

274

275 When applied using Simo and Dachs' [2002] global mean DMS flux estimate
 276 equation (18) produces a CCN estimate of $140 \text{ particles cm}^{-3}$, comparable to CCN
 277 observations of $50\text{-}250 \text{ particles cm}^{-3}$ reported for the Northeast Atlantic Ocean by
 278 Hegg [1994]. Substituting equation (16) into equation (18) then gives:

279

$$\begin{aligned} CCN &= 710 (k_6 P + k_7 Z) - 15 \\ &= k_8 (k_6 P + k_7 Z) - k_9 \end{aligned} \quad (19)$$

281

282 where $k_8 = 710$ and $k_9 = 15$.

283

284 **Cloud condensation nuclei and cloud droplets**

285 Several authors have derived relationships between CCN and cloud droplet number
 286 concentration (N_C) (see Gondwe [2004] for a summary). The empirically measured
 287 relationships reported in the literature take several forms, although they all describe
 288 similar (hyperbolic-like) relationships. In keeping with our heuristic approach, we
 289 have used the simple non-linear relationship between cloud droplet number (N_C) and
 290 the number of CCN measured by Saxena and Menon [1999] over the south-eastern
 291 United States:

292

$$\begin{aligned}
 N_C &= 183 \ln(CCN) - 334 \\
 &= k_{10} \ln(CCN) - k_{11} \quad ,
 \end{aligned}
 \tag{20}$$

294

295 where $k_{10} = 183$ and $k_{11} = 334$. This relationship is similar (differing only slightly in
 296 the coefficients) to the relationship between CCN and the sub-cloud aerosol number
 297 concentration measured over the North Atlantic Ocean by Gultepe and Isaac [1996]. It
 298 also produces estimates of N_C similar to the theoretical derivation of Chuang and
 299 Penner [1995] (when applied with the associated relationship between CCN and
 300 sulphate aerosol mass concentration given by Saxena and Menon [1999]) suggesting
 301 equation (20) is a robust parameterisation. Substituting equation (19) into equation
 302 (20) gives an estimate of the number of cloud droplets that will result from DMS
 303 ventilated into the atmosphere from the ecosystem:

304

$$N_c = k_{10} \ln \left[k_8 (k_6 P + k_7 Z) - k_9 \right] - k_{11}. \quad (21)$$

306

307 **Cloud droplets, albedo and irradiance**

308 Relationships between changes in cloud droplet number and changes in cloud top
 309 albedo (α), assuming constant atmospheric liquid content, have been provided by
 310 several authors [Han, *et al.*, 1998]. These are all similar linear relationships, varying
 311 only in the slope, and the most recent, given by Schwartz and Slingo [1996], and valid
 312 for albedos between 0.28 and 0.72, is used here:

313

$$\begin{aligned} \Delta\alpha &\approx 0.075 \left(\frac{\Delta N}{N_0} \right) \\ &\approx 0.075 \left(\frac{N_c - N_0}{N_0} \right), \end{aligned} \quad (22)$$

315

316 where N_0 is the reference cloud droplet number density and ΔN is the change in cloud
 317 droplet concentration from the reference. The change in cloud top albedo due to
 318 change in the flux of DMS from the ocean to the atmosphere can then be
 319 approximated by substituting equation (21) into equation (22):

320

$$\begin{aligned}
\Delta\alpha &= 0.075 \left\{ \frac{k_{10} \ln \left[k_8 (k_6 P + k_7 Z) - k_9 \right] - k_{11} - N_0}{N_0} \right\} \\
&= k_{12} \left\{ \frac{k_{10} \ln \left[k_8 (k_6 P + k_7 Z) - k_9 \right] - k_{11} - N_0}{N_0} \right\}, \quad (23)
\end{aligned}$$

where $k_{12} = 0.075$.

If it is assumed that the reference number of droplets (N_0) is equated with the steady state of the ecosystem (i.e. the reference number of droplets includes a contribution from the ecosystem at steady state), then N_0 can be described in terms of the steady state values of the *NPZ* model:

$$N_0^* = k_{10} \ln \left[k_8 (k_6 P^* + k_7 Z^*) - k_9 \right] - k_{11}, \quad (24)$$

where P^* and Z^* are defined by equations (6) and (7) respectively. The change in cloud top albedo attributable to a change in DMS production by the ecosystem is then given by:

$$\Delta\alpha = \frac{k_{12}}{N_0^*} \left\{ k_{10} \ln \left[k_8 (k_6 P + k_7 Z) - k_9 \right] - k_{11} - N_0^* \right\}. \quad (25)$$

338 Irradiance and photosynthesis

339 The depth of mixing in the upper ocean influences the average irradiance experienced
 340 by phytoplankton. This also varies latitudinally, seasonally and daily with values of
 341 30m typical of equatorial regions, and 100m typical of high latitudes. The average
 342 mixed layer irradiance at any latitude may be approximated by:

343

$$344 \quad I_A = \frac{I_\theta}{MLD} \int_0^{MLD} e^{-k_L z} dz = \frac{I_\theta}{MLD} \left(\frac{1 - e^{-k_L MLD}}{k_L} \right), \quad (26)$$

345

346 where I_A is the average irradiance over the mixed layer,

347 $I_\theta = I_E \cos(\theta)$ is the incident surface irradiance at latitude θ ,

348 I_E is the incident surface irradiance at the equator,

349 MLD is the depth of the mixed layer,

350 z is depth in the water column in metres, and

351 k_L is the seawater light extinction coefficient (typically 0.04 m^{-1} for
 352 ocean water).

353

354 Zonneveld [1998] derived a photosynthesis-irradiance (PI) curve of the general form:

355

356
$$P_{photo} = \frac{aI_p}{bI_p^2 + I_p + c}, \quad (27)$$

357

358 where P_{photo} denotes photosynthesis, I_p denotes irradiance measured as the average
 359 number of photons per area per time, a represents the maximum rate quantum yield
 360 per photosynthetic unit, b represents the ratio of the cell absorption cross-section to
 361 the specific recovery rate of damaged d -protein, and c represents the excitation
 362 requirements of the cell. Zonneveld observed that other researchers had derived
 363 relationships of the same general form from different premises.

364

365 Phytoplankton acclimate to the average light intensity in which they grow [Zonneveld,
 366 1998], and it is therefore reasonable to assume that phytoplankton throughout the
 367 global oceans are adapted to their local irradiance. The average irradiance experienced
 368 by endemic phytoplankton at any latitude on the globe (I_A) may then be scaled by
 369 their saturating irradiance (I_S , at which phytoplankton photosynthesis is a maximum),
 370 and the effect of variations in irradiance on phytoplankton growth at any latitude may
 371 then be approximated by:

372

373
$$P_{photo} = \frac{aI}{bI^2 + I + c}, \quad (28)$$

374

375 where $I = \frac{I_A}{I_S}$. The maximum value of equation (28) occurs when:

376

377

$$I = \sqrt{\frac{c}{b}}, \quad (29)$$

378

379 and the maximum value of P_{photo} is now by definition achieved when $I = 1$, which
 380 from equation (29) stipulates that $c = b$. The PI relationship may be applied to a
 381 particular phytoplankton species by scaling it to be a function between 0 and 1 that
 382 operates on the maximum phytoplankton growth rate (k_I) of the phytoplankton
 383 species. This condition specifies that $P_{photo}(I = 1) = 1$, and therefore equation (28)
 384 stipulates that $a = 1 + 2b$. The photosynthesis-irradiance relationship may then be
 385 written as a general non-dimensional operator on the maximum phytoplankton growth
 386 rate:

387

388

$$R = \frac{(1 + 2b)I}{bI^2 + I + b}, \quad (30)$$

389

390 where b is now a photo-inhibition parameter that controls the shape of the PI curve
 391 (Figure 2) and is likely to be species dependent. Note that R is relatively insensitive to
 392 b once $b > 1$. As b tends towards infinity R tends to $\frac{2I}{I^2 + 1}$, so when $b = \infty$, $R = 0.385$
 393 at $I = 5$, not appreciably different to its value when for example $b = 10$, when $R =$
 394 0.396.

395

396 The R curves in Figure 2 represent a dimensionless operator on the maximum
 397 phytoplankton growth rate, and equation (2) in the ecosystem model becomes:

398

$$399 \quad \frac{dP}{dt} = k_1 R \left(\frac{N}{N + k_2} \right) P - k_3 PZ, \quad (31)$$

400

401 and the appropriate modification is also made to equation (1):

402

$$403 \quad \frac{dN}{dt} = k_5 Z + k_3 k_4 PZ - k_1 R \left(\frac{N}{N + k_2} \right) P. \quad (32)$$

404

405 The usual condition when the climate is unperturbed (by anthropogenic or biogenic

406 activities) has $I = 1$ (as we have previously assumed $I = \frac{I_A}{I_S}$ in equation (28)). Any

407 perturbations to this steady state due to ecosystem feedbacks such as described by

408 equation (23) will affect the irradiance experienced by the phytoplankton, and can be

409 modelled by replacing the irradiance in equation (30) with the term $1 - \Delta\alpha$,

410 representing the change in irradiance due to changes in albedo:

411

$$R = \frac{(1+2b)(1-\Delta\alpha)}{b(1-\Delta\alpha)^2 + (1-\Delta\alpha) + b}. \quad (33)$$

413

414 This feedback effect may be written in terms of P and Z by substituting for $\Delta\alpha$ from
 415 equation (25) giving an expression describing the effect on a marine planktonic
 416 ecosystem of its production of DMS:

417

$$R = \frac{(1+2k_{13}) \left(1 - \frac{k_{12}}{N_0} \left\{ k_{10} \ln [k_8 (k_6 P + k_7 Z) - k_9] - k_{11} - N_0 \right\} \right)}{k_{13} \left(1 - \frac{k_{12}}{N_0} \left\{ k_{10} \ln [k_8 (k_6 P + k_7 Z) - k_9] - k_{11} - N_0 \right\} \right)^2 + \left(1 - \frac{k_{12}}{N_0} \left\{ k_{10} \ln [k_8 (k_6 P + k_7 Z) - k_9] - k_{11} - N_0 \right\} \right) + k_{13}}, \quad (34)$$

419

420 where k_{13} has been substituted for b .

421

422 The feedback model

423 A global biogeochemical feedback model incorporating the effects of DMS produced
 424 by marine ecosystems on climate and irradiance, and therefore their own environment,
 425 can now be written by incorporating equation (34) into the NPZ model (equations (1)
 426 - (3)):

427

$$\frac{dN}{dt} = k_5 Z + k_3 k_4 P Z - R k_1 \left(\frac{N}{N + k_2} \right) P, \quad (35)$$

428

$$\frac{dP}{dt} = Rk_1 \left(\frac{N}{N + k_2} \right) P - k_3 PZ, \quad (36)$$

$$\frac{dZ}{dt} = k_3 (1 - k_4) PZ - k_5 Z, \quad (37)$$

431

432 where R is given by equation (34). The values of the additional parameters ($k_6 - k_{13}$)
 433 used for the feedback model simulations are listed in Table 2. The nature of the
 434 scaling described in the model formulation ensures that the steady states of the simple
 435 and feedback models are identical (i.e. at steady state $R = 1$).

436

437 **The time-lagged feedback model**

438 The time-lagged feedback model was developed by including a time lag (τ) in the
 439 feedback term:

440

$$R = \frac{(1 + 2k_{13}) \left(1 - \frac{k_{12}}{N_0} \left\{ k_{10} \ln \left[k_8 (k_6 P(t - \tau) + k_7 Z(t - \tau)) - k_9 \right] - k_{11} - N_0 \right\} \right)}{k_{13} \left(1 - \frac{k_{12}}{N_0} \left\{ k_{10} \ln \left[k_8 (k_6 P(t - \tau) + k_7 Z(t - \tau)) - k_9 \right] - k_{11} - N_0 \right\} \right)^2 + \left(1 - \frac{k_{12}}{N_0} \left\{ k_{10} \ln \left[k_8 (k_6 P(t - \tau) + k_7 Z(t - \tau)) - k_9 \right] - k_{11} - N_0 \right\} \right) + k_{13}} \quad , \quad (38)$$

442

443 so that phytoplankton growing at time (t) experience light conditions determined by
 444 *DMS* emissions generated by the P and Z populations at some earlier time ($t - \tau$).

445

METHODS

NPZ-DMS model validation

Often the only model validation available for ecosystem models is to test whether a model can reproduce observed data [Franks, 2002], and the *NPZ-DMS* model (equations (1) - (5)) used in this analysis was evaluated against this criteria in a study region in the Southern Ocean off the eastern Antarctic coast (60-65° S, 125-140° E). The model was formulated as a seasonally-forced, depth-averaged model for the validation, where the state variables were averaged over the depth of the surface ocean mixed layer and the model was forced with climatologies of environmental data. The forcings comprised ocean mixed layer depth (*MLD*), sea surface temperature (*SST*), photosynthetically active radiation (*PAR*) and sea surface wind speed (*WIN*), and were sourced from the World Oceanographic Atlas, and from the Pathfinder, SeaWiFS and SeaWinds satellite sensors respectively. Time series of the forcings are shown in Figure 3.

Annual climatologies of ocean surface chlorophyll and DMS concentrations for the study region were developed from SeaWiFS satellite data and the Kettle et al [1999] database respectively. Model *P* values in mg N m⁻³ were converted to mg Chl *a* m⁻³ assuming a Redfield C:N ratio of 5.7 (by weight) and a typical carbon : chlorophyll ratio of 50 [Walsh, et al., 2001] for the comparison. A genetic algorithm, an efficient non-linear optimisation technique that does not require any derivative information [Holland, 1975; Mitchell, 1997], was used to fit the model to observed data by

minimising the squared error between the model predictions and the observed data. A parameter set was derived for the forced *NPZ* model so that the *P* population reproduced the surface ocean chlorophyll concentrations climatology. The *NPZ-DMS* model was then fitted to the DMS climatology

Seasonal forcing is implemented in the depth-averaged form of the *NPZ-DMS* model by replacing k_1 in equations (1) and (2) with k'_1 , where:

$$k'_1 = R_L R_T k_1, \quad (39)$$

where R_T is the temperature limitation of the phytoplankton growth rate and R_L is the light limitation of the phytoplankton growth rate. The light limitation factor R_L is used to explicitly represent the measured seasonal forcing of light in driving chlorophyll and hence DMS dynamics for the model validation (where it must reproduce seasonal variations in chlorophyll and DMS). In contrast the light factor R (equation (34)) represents the feedback light limitation due to changes in albedo, and is free of seasonal effects in an unforced model.

Laboratory studies of phytoplankton have revealed a dependence of phytoplankton growth rates on temperature [Eppley, 1972; Goldman and Carter, 1974]. The temperature dependence of phytoplankton growth used in the model validation was estimated by Eppley [1972] to be:

491

$$R_T = e^{0.063(T - T_{max})}, \quad (40)$$

493

494 where T is the ambient temperature ($^{\circ}\text{C}$) and T_{max} is the maximum annual temperature.

495 R_T was forced with the climatology of SST (Figure 3a) for the validation.

496

497 As noted previously, the average irradiance in the ocean mixed layer (I_A) can be
 498 approximated by equation (26). For the purposes of the model validation, a form of
 499 light limitation on the phytoplankton growth rate proposed by Walsh et al. [2001] was
 500 used:

501

$$R_L = \frac{I_A}{I_S} e^{\left(1 - \frac{I_A}{I_S}\right)}, \quad (41)$$

503

504 where I_S the phytoplankton saturating irradiance (assumed to be 35 W m^{-2} for this
 505 analysis ([Walsh, et al., 2001], and references cited therein). R_L was forced with the
 506 climatologies of MLD and PAR (Figure 3 b and c) for the validation.

507

508 The *DMS* model (equations (4) - (5)) was modified for the validation exercise by
 509 including a term parameterising the transfer of DMS from the ocean to the atmosphere
 510 identical to that used in the feedback model (equations (12) - (15)). However, the *DMS*

sea-air transfer velocity (k_{tr}) was forced by the climatologies of SST and WIN (Figure 3 a and d) for the validation exercise.

Perturbation analysis

The eigenvalues of the *NPZ* model are easily derived and informative, however, the eigenvalues of the biofeedback model are not. The effect of including the feedback on the model dynamics was therefore evaluated by numerically calculating the time required for each model to return to steady state after a perturbation. The return time of a system provides a useful measure of its resilience as defined by DeAngelis [1980], to which it is inversely proportional.

The models are reactive in the sense of Neubert and Caswell [1997] in that they respond to almost all perturbations by initially amplifying the magnitude of the perturbation, after which the perturbation exponentially decays as the model returns to its steady state. A simple metric measuring the closeness of the model trajectory to steady state was not therefore possible, and the time required for the whole of the model's trajectory to be confined within a circle of 0.05% of the area of the model's state space was therefore used. The accuracy of this metric is determined by the period of oscillation of the system and is approximately controlled by the imaginary part of the linearised non-feedback system's eigenvalue given by:

$$Err \approx \pm \frac{2\pi}{\text{Im}(\lambda)} = \pm 4\pi^{-2} \sqrt{\left[k_1 \left(\frac{k_2}{(N^* + k_2)^2} \right) P^* \right]^2 - 4P^* \left[k_3 k_4 Z^* - k_1 \left(\frac{N^*}{N^* + k_2} \right) \right] \left[k_3 - k_1 \left(\frac{k_2}{(N^* + k_2)^2} \right) P^* \right]}. \quad (42)$$

534

535 Return time surfaces were calculated for each of the models to demonstrate the effect
 536 of the inclusion of the feedback process on the model resilience. These surfaces were
 537 generated by perturbing each model 400 times using combinations of perturbations of
 538 -90 to +100 % (in steps of 10%) of the steady state values of P and Z (equations (6)
 539 and (7) respectively). The models were integrated forward in time using the perturbed
 540 steady state values as initial conditions for the integration. Return time surfaces were
 541 generated for each model and normalised by the largest return time of the non-
 542 feedback model. A difference surface describing the effect of the feedback was also
 543 calculated. Although the return time surfaces appear quite smooth, the accuracy
 544 constraints of the metric used to build them meant that the difference surface was not
 545 smooth. The difference surface was therefore smoothed for display; however, the
 546 smoothing did not change the fundamental characteristics of the surface.

547

548 Sensitivity analysis

549 A sensitivity analysis of the biofeedback model was undertaken to ascertain the most
 550 important parameters and processes in the model and to evaluate its behaviour. The
 551 New Morris Method, an efficient second order screening method,[*Campolongo and*
 552 *Braddock, 1999; Cropp and Braddock, 2002*] was used for the sensitivity analysis.
 553 The sensitivity analysis measured the difference in return time surfaces of the models.
 554 This analysis quantified the sensitivity of the feedback effect to the model parameters,

and is an important part of the analysis because, as was evident in the development of the feedback model, the empirical relationships used have poorly known parameter values. Two sensitivity metrics were calculated; the sum of the differences between the return times of the models for each perturbation:

$$F = \sum_{i=1}^{20} \sum_{j=1}^{20} (R_{N:i,j} - R_{F:i,j}), \quad (43)$$

where $R_{N:i,j}$ is the return time for the *NPZ* model from perturbation (i, j) and $R_{F:i,j}$ is the equivalent return time of the feedback model, and the normalised equivalent:

$$F_N = \sum_{i=1}^{20} \sum_{j=1}^{20} \left(\frac{R_{N:i,j} - R_{F:i,j}}{R_{N:i,j}} \right). \quad (44)$$

The sensitivity analysis was implemented to examine the influence of the twenty-two parameters of the feedback model ($k_1 - k_5, k_8 - k_{13}, m_1 - m_7$ and γ , see Tables 1 and 2) and was implemented using parameter ranges of $\pm 25\%$ of the values listed in Tables 1 (G99) and 2. The analysis made 48,400 comparisons of return time surfaces and required 3.87×10^7 model evaluations.

573 **Time lag analysis**

574 The inclusion of the feedback time lag analysis recognises that a delay between a
575 phytoplankton bloom and increased DMS concentration in the water has been
576 observed [Turner, *et al.*, 2004; Turner, *et al.*, 1996] and that time lags between the
577 transfer of DMS to the atmosphere and modification of irradiance are likely to be
578 significant. As none of the relationships used to build the feedback model are reported
579 with associated time scales, the feedback model by default includes the assumption
580 that the atmospheric processes of the feedback occur instantaneously. Clearly this
581 assumption is untenable, and hence we have also examined the effect of time lags in
582 the feedback process on our simulation results.

583

584 The effect of time lags in the feedback processes was examined by modifying the
585 feedback model to include a time lag into the feedback R that operates on the P
586 growth term (see feedback model description above). However, there is little
587 conclusive evidence indicating the magnitude of the time lags involved in these
588 processes [Ayers and Gillett, 2000]. In situ correlations of ocean-atmosphere DMS
589 flux, atmospheric DMS concentration and condensation nuclei [Andreae, *et al.*, 1995]
590 have suggested that some pathways may be as short as six hours [Lin and Chameides,
591 1993]. In the absence of definitive evidence to the contrary, a reasonable first
592 approximation is to commence the time lags at zero and increase them until
593 interesting model behaviour appears exhausted.

594

A numerical analysis of the difference between the return time surfaces of the *NPZ* model and the time-lagged feedback model was conducted for time lags between 0 and 15 days. The perturbation metric selected ensured that the model was integrated for approximately 1,500 days, about a hundred times longer than the largest time lag, ensuring the discontinuities related to the initial conditions did not affect the return times. Return time surfaces were calculated for 60 time lags between 0.25 and 15 days in increments of 0.25 days. These return time surfaces were each subtracted from the return time surface of the *NPZ* model and the resulting difference surface was summed to give a scalar metric of the feedback effect for each time lag. This metric was scaled by the sum of the return time surface of the equivalent *NPZ* model to provide a proportional measure of the feedback effect. This enabled comparison of the feedback effects between different parameterisations as they have very different magnitude return time surfaces.

The ecosystem model contains three rate parameters: the phytoplankton maximum growth rate (k_1), the rate of zooplankton grazing on phytoplankton (k_3) and the rate of zooplankton mortality (k_5). Previous sensitivity analysis of the major determinants of DMS flux to the atmosphere [Cropp, *et al.*, 2004] have identified the importance of k_1 and k_3 . These results are confirmed by the importance of these parameters to the feedback effect examined in this research (see below), which also identified k_5 as an important determinant of the magnitude of the feedback effect. The time lag analysis was therefore conducted for the basic parameter set and also for parameter sets with doubled k_1 , doubled k_3 or doubled k_5 .

RESULTS

NPZ-DMS model validation

The P concentration predicted by the depth-averaged *NPZ* model for the best-fit parameter set derived by the genetic algorithm is shown in Figure 4 (a) with the SeaWiFS chlorophyll data. Similarly, the best fit to the interpolated DMS data derived by Kettle et al (1999) is shown in Figure 4 (b). (The environmental forcings used in the simulation are shown in Figure 3.) The parameter values used to generate the predictions in Figure 4 are listed in Table 1 (GA). The model produces a prediction that matches the observed data well, suggesting that the *NPZ* model is a valid representation of the generic plankton seasonal dynamics in the region of the Southern Ocean off the east Antarctic coast, and similarly that the *DMS* model is a valid representation of the seasonal DMS dynamics in this region.

Return time surfaces

The return time surface for the simple *NPZ* model without biogenic feedback on its environment is shown in Figure 5. This reveals that the model is more sensitive to perturbations in Z than to perturbations in P , in fact perturbations in P have almost no effect on the return time of the system if Z has suffered a large perturbation in either direction. This return time surface is characteristic of the *NPZ* model and is robust to substantial (but not all) parameter variations

The return time surface for the feedback model is shown in Figure 6. This return time surface is very similar to the surface for the *NPZ* model without biogenic feedback, the principal difference being that the feedback model returns to equilibrium more rapidly than the non-feedback model.

The surface representing the difference in return times to equilibrium for each of the models is shown in Figure 7. This surface clearly demonstrates that the inclusion of the biogenic feedback on its environment has endowed the *NPZ* model with increased resilience. The time the system requires to recover its equilibrium state after perturbations of any magnitude in any direction is substantially reduced by the inclusion of the feedback process. Similarly to the models' return time surfaces, the magnitude of the *P* perturbation does not have a substantial effect on the difference between the two return time surfaces. The magnitude by which the resilience is enhanced by the feedback process appears to be largely determined by the *Z* perturbation. For each *Z* perturbation, the *P* perturbations appear to have little influence on the return time of the system.

This increased resilience is an interesting result as the feedback (*R*) is formulated so that at steady state $R = 1$ (its maximum possible value) and the resilience of the feedback model is identical to that of the non-feedback model. Therefore including the feedback into the model cannot increase the resilience (as defined by equation (10)) of the model. The effect of the feedback when the *P* and *Z* populations are greater than their steady state values, the effect of the feedback is intuitive – it serves to reduce the irradiance the *P* receive and slows their growth rate, helping them to

decline back to the steady state value. The effect on the system when the P and Z populations are smaller than their steady state values is not so intuitive – the P growth rate is again slowed, but it is not clear how this helps them to achieve their steady state values more rapidly.

Sensitivity analysis

All of the 48,400 comparisons of un-lagged return time surfaces indicated that the biofeedback model returned to steady state more rapidly than the non-feedback model, that is that the addition of the biogenic feedback always reduced the time taken for the system to recover from perturbation. This result indicates that the stabilising effect of adding the instantaneous feedback to the ecosystem model is robust to substantial parameter variations of both ecosystem and atmospheric processes.

The sensitivity analysis indicated that the model return times after perturbation were sensitive to both biological parameters describing the ecosystem and physical parameters describing the atmospheric processes. Both the raw and normalised metrics used to measure the sensitivities (equations (43) and (44)) revealed almost the same sensitivities to the parameters. Only one of these, the raw sensitivities, are therefore presented, with Figure 8 describing the first order effects and Figure 9 the second order or two-parameter interaction effects. Two of the most influential parameters were k_{10} and k_{11} , describing the cloud droplet to CCN relationship (equation (20)), followed by k_3 , the Z grazing rate, k_1 , the P growth rate, and k_5 , the Z

mortality rate. It is apparent from equation (10) that these ecosystem rate parameters also have significant impacts on the model resilience.

Combinations of these parameters were also important in the two-factor effects shown in Figure 9. While the two most influential parameter interactions, k_{10}, k_{11} and k_1, k_3 are atmospheric and biological pairs respectively, the next most important pair k_3, k_{10} and several other important pairs involving k_1, k_5, k_{11} and k_{12} reveal that interactions between the marine ecological processes and the atmospheric physical and chemical processes are important determinants of the magnitude of the feedback effect.

The three important results from the sensitivity analysis are therefore that the consequences of the feedback for the ecosystem depend on both biological and atmospheric properties, that an instantaneous feedback always acts to stabilise the ecosystem dynamics, and that it is critically important to the determination of the actual magnitude of this feedback process that both the atmospheric and biological components of the process noted above are better quantified by field measurements.

Time lag analysis

The time lag analysis reveals that the magnitude and direction of the feedback effect is dependent on the time lag between the ecosystem dynamics and the change in irradiance experienced by the phytoplankton as a result of the atmospheric feedback (Figure 10). The solid line in Figure 10 shows the effect of feedback time lags on the return time of the feedback model for the basic parameter set. This reveals that the

initial stabilising effect (by which is meant that the resilience the system is increased) of the feedback, where it reduces the return time of the model by about 6%, rapidly declines once a lag of two days is introduced into the feedback. The feedback has no effect on the resilience of the model if the feedback is lagged by three days, and for lags of three to six days, the feedback destabilises (reduces the resilience of) the model, increasing the return time by up to about 6%.

Time lags of six to ten days once again increase the resilience of the feedback model (by up to about 2%). This recovery of the stabilising effect of the feedback, with a maximum in the second stabilising region at a time lag of about seven days is especially interesting given that ocean in situ ecosystem-scale experiments that stimulated phytoplankton blooms measured a peak in DMS aqueous concentrations about a week after the bloom [Turner, *et al.*, 1996]. Time lags greater than ten days again result in destabilisation of the feedback model. It should be noted, however, that while the lagged feedback sometimes makes this model less resilient, causing it to take up to 10% longer to reach steady state than the non-feedback model (Figure 10), it never reduces the resilience to zero (i.e. the system always returns to the steady state). For the realistic (i.e. based on measured values) parameter values used (Tables 1 and 2) both models are always stable – only the resilience, and hence the time taken to return to steady state after a perturbation, is affected by the feedback.

The time lag analysis also reveals the influence that the properties of the biota in the ecosystem can have on the feedback process. The simulation with k_I doubled (Figure 10, dotted line) reveals that the increased phytoplankton growth rate both greatly

reduces the time lags for which the feedback model is initially more resilient (two days rather than three) and also reduces the magnitude of the stabilising or destabilising effects (a 4% reduction in return time). Similarly the recovery of the stabilising effect occurs earlier (around four days), but lasts for only two days and produces only a 1% reduction in return time.

The simulation with doubled zooplankton grazing (k_3) reveals quite the opposite effect (Figure 10, dashed line). In this case the increase in resilience for lags of up to three days is substantially increased with return times being up to 12.5% shorter than the non-feedback model. The region of reduced resilience is smaller and the stabilising effect is recovered much sooner at about 4 days. This variant also recovers resilience to a greater extent than the base parameter feedback model, with reductions in return time of up to 6%. The reduction in resilience that subsequently occurs is again very similar to the base parameter feedback model. The distinctive feature of the increased zooplankton grazing model is that its region of reduced resilience in Figure 10 is much smaller (in magnitude and duration) compared to both other parameter options.

An increase in the rate of zooplankton mortality (k_5) also reduces the stabilising effect of the feedback, reducing its initial increase in resilience to about 2%, but providing the largest reduction in resilience of over 7.5% at about day three. This variant also reduces the duration of the initial stabilising feedback to less than 2 days. The feedback again produces increased resilience after a time lag of 5 days, and remains so up to about 8 days, but only marginally (less than 1% reduction in return time). However, subtle effects that operate for long periods of time may still be influential.

758

759 An interesting aspect of the response of the feedback effect to these parameter
760 variations is that the feedback process provides the ecosystem with a mechanism to
761 recover resilience if the attributes of the biota change to reduce it. Increases in the
762 phytoplankton growth rate (k_1) and the zooplankton mortality rate (k_5) reduce the
763 stabilising effect of the feedback, but it is apparent from equation (10) that they
764 simultaneously must increase the resilience of the ecosystem. Similarly, increases in
765 the zooplankton grazing rate (k_3) reduce the resilience of the ecosystem, but increase
766 the stabilising effect of the feedback. Of these two processes, the ecosystem resilience
767 is the principal determinant of the response to perturbation; the feedback system with
768 increased phytoplankton growth rate has shorter return times than the system with
769 increased zooplankton grazing, even though the latter provides a much greater
770 stabilising feedback effect. An interesting heuristic gained from this analysis is that
771 the feedback process provides a “resilience insurance policy” in that changes in the
772 properties of the ecosystem that tend to reduce its resilience are countered by an
773 increase in resilience delivered by the feedback, and *vice versa*.

774

775 **DISCUSSION**

776 This research essentially generates a hypothesis that the DMS feedback process may
777 serve to stabilise (that is, increase the resilience of) some ecosystems. The
778 geographical extent to which this hypothesis may apply is as yet unclear, as the *NPZ-*
779 *DMS* model is not able to reproduce chlorophyll and DMS dynamics over the whole
780 of the global oceans. The stability attributes of the *NPZ* and *DMS* models suggest that
781 the model will be best able to reproduce observed dynamics in regions of the oceans

782 where the chlorophyll seasonal variation closely follows the physical forcings of
783 irradiance, temperature and mixed layer depth, and the DMS concentration is closely
784 coupled to the chlorophyll signal. This is generally the situation in high latitudes
785 where the forcings are very strong and drive the plankton dynamics, but is not the
786 case in many parts of the oceans. In the equatorial oceans, for example, chlorophyll is
787 often out of phase with the physical forcings, and this model cannot reproduce the
788 temporal chlorophyll dynamics in these regions. Similarly, the model cannot
789 reproduce DMS dynamics if there is a substantial time lag between the chlorophyll
790 maximum and the DMS maximum, for example the “summer paradox” observed at
791 some mid and low latitude locations [*Simo and Pedros-Alio, 1999*].

792
793 These caveats constrain the spatial range where the model can competently reproduce
794 measured data, but do not necessarily degrade the heuristic value of the model. Work
795 in progress by the authors suggests that the dynamics of the *NPZ* model used in this
796 analysis is a subset of a slightly more complicated *NPZ* model that can fit chlorophyll
797 dynamics that are out of phase with their physical forcings. (The dynamics of this
798 slightly more complicated model are such that at this stage it does not have the
799 heuristic value of the model we have used.) Similarly, Vallina et al (in preparation)
800 show that the DMS summer paradox may be explained by including an extra DMS
801 source term, reflecting the exudation of DMS by phytoplankton under UV stress, that
802 effectively loosens the coupling between the ecosystem and chemical models.

803
804 Although there are no assumptions in the model that intrinsically limit its application
805 to any part of the globe, the validation exercise suggests that inferences and

hypotheses drawn from this exercise may only be directly relevant to high latitude regions. Never-the-less, this research contributes a new dimension to the DMS hypothesis of Charlson et al [1987] in particular, and to the field of Earth System Science in general. Our results show that an instantaneous biogenic feedback on the planktonic ecosystem model's irradiance environment can make the model more resilient to perturbation. The time that the system requires to return to its equilibrium state after any perturbation is therefore reduced by the addition of the instantaneous feedback.

The stabilising effect of the feedback is largest for perturbations that reduce the *Z* population. This is an interesting result, as recent work on model *NPZ* ecosystems [Cropp and Norbury, 2007] suggests that extinction processes in climate change scenarios will progress sequentially commencing with the extinction of *Z*. The presence of the feedback mechanism in the model therefore provides the system with an additional safeguard against extinction: the smaller the *Z* population gets, the stronger the feedback restoring the system to its equilibrium state becomes. This suggests the heuristic that the stability properties engendered by the feedback act most strongly in the manner required to both save the species and to maintain the ecosystem in its original form.

The sensitivity analysis revealed two important properties of the instantaneous feedback effect: that it always serves to increase the resilience of the model, and that it is a function of both the biological and physical-chemical factors involved in the process. Two important parameters controlling the magnitude of the feedback effect

are the slope (k_{10}) and intercept (k_{11}) values of the regression relating the number of cloud droplets in the atmosphere to the number of cloud condensing nuclei in the atmosphere. The second most influential parameter is the rate of zooplankton grazing on phytoplankton (k_3). This parameter was identified by Cropp et al. [2004] as the most important determinant of annual integrated DMS flux to the atmosphere.

The time lag analysis revealed that the magnitude of the feedback effect, and whether it increases or reduces the resilience of the system, depends on the time lags associated with the feedback processes. This analysis revealed that the feedback could make the system more sensitive to perturbation if the effect of the feedback took several days to impact on the ecosystem. However, the most interesting and perhaps important result of the time lag analysis is that for time lags of about 7-10 days the feedback effect again increases the resilience of the ecosystem; that is the feedback causes the ecosystem to return to its equilibrium state more rapidly than if the feedback was not present. The time lags that lead to this recovered stabilising effect correspond with time lags observed in *in situ* ocean ecosystem experiments between phytoplankton blooms and subsequent maximum aqueous DMS concentrations [Turner, et al., 2004; Turner, et al., 1996]. This analysis also identified an important modification to the results of the sensitivity analysis that also serves to further reflect the importance of k_1 , k_3 and k_5 on the stabilising effects of the feedback, and suggest that the timing and magnitude of the feedback effects will vary between ecosystems.

The time-lagged feedback model we have used does not account for spatial factors, and the rapid advection rates in the atmosphere compared to the ocean suggest that

only in periods of calm weather would the atmospheric effects generated by a phytoplankton bloom affect the originating population. However, as the atmospheric processes involved in the feedback may be rapid [Hamilton and Lenton, 1998], for some pathways perhaps as short as six hours [Lin and Chameides, 1993], couplings on local and regional scales may be close.

The influence of the feedback on the ecosystem model's stability characteristics leads to the interesting hypothesis that the DMS feedback cycle may not be just an artefact of plankton biochemistry, but an intrinsic component of marine planktonic ecosystems. Such a hypothesis would suggest that marine phytoplankton gained a benefit from the production of DMSP as it initiated processes that stabilised the plankton ecosystem and consequently buffered the population from the effects of perturbations. Many authors, perhaps beginning with Dunbar [1960], have pointed to the susceptibility of populations to extinction due to stochastic events when at the nadir of extended "boom and bust" cycles that many low resilience systems experience after perturbation.

Such a hypothesis would require the invocation of group selection, a concept extensively attacked in the 1970's (see for example Dawkins [1976]). While it is not the intent of this research to debate the merits of group selection, we note that group selection is attracting new interest in Western evolutionary debate [Borello, 2005] and that ecosystem evolution is an accepted paradigm in Russian science [Lekiavichius, 2003]. As noted by Loreau et al. [2004] "Species traits and their evolution are

ultimately constrained by ecosystem processes, just as ecosystem properties are constrained by the ecological and evolutionary history of interacting species.”

Our results clearly show that the influence of the feedback is dependent on the time lags in the ocean and atmospheric processes, and on the characteristics of the ecosystem. Our theoretical analysis suggests that clarification of the time scales of the feedback processes, and better knowledge of all the parameters in the model, would be a very useful contribution to the whole DMS hypothesis.

CONCLUSION

This research has modelled a process whereby production of a precursor compound by individual phytoplankton, coupled with subsequent biological processing by co-constituents of a plankton ecosystem, results in a product that modifies the properties of the atmosphere. The changes in albedo we have modelled subsequently influence the irradiance experienced by the phytoplankton that initiated the process. Our results demonstrate that this change in environment modifies the population dynamics of the phytoplankton and may make the ecosystem to which they belong more resilient to perturbation. Our results also suggest that the magnitude and timing of the feedback effects are highly dependent on the characteristics the ecosystems and the atmospheric processes involved. Although the feedback effect can reduce the resilience of the ecosystem when time lags are introduced, the feedback increases the resilience of the system at time lags similar to those observed in real ocean plankton ecosystems.

900 The coherence of these theoretical and observed relationships then raises the question
901 of whether the phytoplankton – DMS – cloud relationship reflects a fortuitous artefact
902 of phytoplankton metabolism or an evolved property that improves phytoplankton
903 fitness by stabilising the dynamics of their ecosystem. Because the benefits derived by
904 an individual phytoplankton from producing DMSP are accrued by the group, and
905 perhaps even the ecosystem as a whole, the option that phytoplankton have evolved
906 the ability to influence the properties of the atmosphere invokes group-level selection.
907 Hamilton and Lenton [1998] have argued just such a case for these systems, although
908 in support of a different hypothesis. The stabilising effects observed in this study,
909 although subtle, would act over long time scales and may therefore have profound
910 implications for the characteristics of the ecosystems that generate them.

911
912 Our research indicates a pressing need for clarification of the time scales of the
913 atmospheric processes involved in the DMS feedback. Although the feedback effects
914 we have documented appear robust to substantial parameter variations the
915 implications of the feedback process for marine plankton ecosystems may not be fully
916 assessed, nor our hypothesis refined, until these details are elucidated. If the time lags
917 observed in real systems do indeed correspond with the time lags that stabilise our
918 model ecosystem, then such further research may yield rich results relating to
919 fundamental properties of living systems.

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REFERENCES

- Andreae, M. O., W. Elbert, and S. J. de Mora (1995), Biogenic sulphur emissions and aerosols over the tropical South Atlantic. 3. Atmospheric dimethylsulphide, aerosols and cloud condensation nuclei., *Journal of Geophysical Research*, 100, 11,335-311,356.
- Ayers, G. P., and R. W. Gillett (2000), DMS and its oxidation products in the remote marine atmosphere: implications for climate and atmospheric chemistry., *Journal of Sea Research*, 43, 275-286.
- Borello, M. E. (2005), The rise, fall and resurrection of group selection., *Endeavour*, 29, 43-47.

- 948 Campolongo, F., and R. Braddock (1999), The use of graph theory in the sensitivity
 949 analysis of the model output: a second order screening method., *Reliability*
 950 *Engineering and System Safety*, 64.
- 951 Charlson, R. J., J. E. Lovelock, M. O. Andreae, and S. G. Warren (1987), Oceanic
 952 phytoplankton, atmospheric sulphur, cloud albedo and climate, *Nature*, 326, 655-661.
- 953 Chuang, C. C., and J. E. Penner (1995), Effects of anthropogenic sulphate on cloud
 954 drop nucleation and optical properties., *Tellus*, 47, 566-577.
- 955 Cropp, R. A., and R. D. Braddock (2002), The New Morris Method: an efficient
 956 second-order screening method., *Reliability Engineering and System Safety*, 78, 77-
 957 83.
- 958 Cropp, R. A., and A. J. Gabric (2002), Ecosystem adaptation: do ecosystems
 959 maximise resilience?, *Ecology*, 83, 2019-2026.
- 960 Cropp, R. A., and J. Norbury (2007), Investigations into a plankton population model:
 961 mortality and its importance in climate change scenarios., *Ecological Modelling*. 201:
 962 97-117.
- 963 Cropp, R. A., J. Norbury, A. Gabric, and R. Braddock (2004), Modeling
 964 dimethylsulphide production in the upper ocean, *Global Biogeochemical Cycles*, 18,
 965 doi:10.1029/2003GB002126.
- 966 Dawkins, R. (1976), *The Selfish Gene*, Oxford University Press, Oxford.
- 967 DeAngelis, D. L. (1980), Energy flow, nutrient cycling and ecosystem resilience,
 968 *Ecology*, 61, 764-771.
- 969 Dunbar, M. J. (1960), The evolution of stability in marine environments: natural
 970 selection at the level of the ecosystem., *The American Naturalist*, 94, 129-136.
- 971 Eppley, R. W. (1972), Temperature and phytoplankton growth in the sea., *Fishery*
 972 *Bulletin*, 70, 1063-1085.
- 973 Franks, P. J. S. (2002), NPZ models of plankton dynamics: their construction,
 974 coupling to physics, and application, *Journal of Oceanography*, 58, 379-387.
- 975 Gabric, A. J., P. A. Matrai, and M. Vernet (1999), Modelling the production and
 976 cycling of dimethylsulphide during the vernal bloom in the Barents Sea, *Tellus*, 51B,
 977 919-937.
- 978 Gabric, A. J., N. Murray, L. Stone, and M. Kohl (1993), Modeling the production of
 979 dimethylsulfide during a phytoplankton bloom, *Journal of Geophysical Research*, 98,
 980 22,805-822,816.
- 981 Goldman, J. C., and E. L. Carter (1974), A kinetic approach to the effect of
 982 temperature on algal growth., *Limnology and Oceanography*, 19, 756-766.
- 983 Gondwe, M. (2004), Quantifying the role of marine phytoplankton in the present day
 984 climate system., 129 pp, University of Groningen.
- 985 Gultepe, I., and G. A. Isaac (1996), The relationship between cloud droplet and
 986 aerosol number concentrations for climate models., *International Journal of*
 987 *Climatology*, 16, 941-946.
- 988 Hamilton, W. D., and T. M. Lenton (1998), Spora and Gaia: how microbes fly with
 989 their clouds., *Ethology, Ecology and Evolution*, 10.

- 990 Han, Q., W. B. Rossow, J. Chou, and R. M. Welch (1998), Global survey of the
 991 relationships of cloud albedo and liquid water path with droplet size using ISCCP.,
 992 *Journal of Climate*, 11, 1516-1528.
- 993 Hegg, D. A. (1994), Cloud condensation nucleus - sulfate mass relationship and cloud
 994 albedo, *Journal of Geophysical Research*, 99, 25,903-925,907.
- 995 Holland, J. H. (1975), *Adaptation in natural and artificial systems.*, 211 pp.,
 996 University of Michigan Press, Ann Arbour.
- 997 Johnson, L. (1990), The thermodynamics of ecosystems., in *The Handbook of*
 998 *Environmental Chemistry 1E: The Natural Environment and the Biogeochemical*
 999 *Cycles.*, edited by O. Hutzinger, pp. 1-47, Springer-Verlag, Berlin.
- 1000 Kettle, A. J., and others (1999), A global database of sea surface dimethylsulfide
 1001 (DMS) measurements and a procedure to predict sea surface DMS as a function of
 1002 latitude, longitude and month., *Global Biogeochemical Cycles*, 13, 399-444.
- 1003 Kiene, R. P., L. J. Linn, and J. A. Bruton (2000), New and important roles for DMSP
 1004 in marine microbial communities., *Journal of Sea Research*, 43, 209-224.
- 1005 Lawrence, M. G. (1993), An empirical analysis of the strength of the phytoplankton-
 1006 dimethylsulfide-cloud-climate feedback cycle., *Journal of Geophysical Research*, 98,
 1007 20,663-620,673.
- 1008 Laws, E. A. (2003), Partitioning of microbial biomass in pelagic aquatic communities:
 1009 maximum resiliency as a food web organizing construct., *Aquatic Microbial Ecology*,
 1010 32, 1-10.
- 1011 Lekivichius, E. (2003), Ecosystem evolution: main stages and potential
 1012 mechanisms., *Zh Obshch Biol*, 64, 371-388.
- 1013 Lin, X., and W. L. Chameides (1993), CCN formation from DMS oxidation without
 1014 SO₂ acting as an intermediate., *Geophysical Research Letters*, 20, 579-582.
- 1015 Liss, P. S., and L. Merlivat (1986), Air-sea gas exchange rates: Introduction and
 1016 synthesis., in *The Role of Air-sea Exchange in Geochemical Cycling.*, edited by P.
 1017 Baut-Menard, pp. 113-127, Reidel, Hingham.
- 1018 Loreau, M., C. de Mazancourt, and R. D. Holt (2004), Ecosystem evolution and
 1019 conservation., in *Evolutionary Conservation Biology.*, edited by R. Ferriere, et al., pp.
 1020 327-343, Cambridge University Press, Cambridge, UK.
- 1021 Mitchell, M. (1997), *An introduction to genetic algorithms.*, 208 pp., MIT Press,
 1022 Cambridge, Massachusetts.
- 1023 Moloney, C. L., M. O. Bergh, J. G. Field, and R. C. Newell (1986), The effect of
 1024 sedimentation and microbial nitrogen regeneration in a plankton community: a
 1025 simulation investigation, *Journal of Plankton Research*, 8, 427-445.
- 1026 Neubert, M. G., and H. Caswell (1997), Alternatives To Resilience For Measuring
 1027 The Responses Of Ecological Systems To Perturbations., *Ecology*, 78, 653-665.
- 1028 Nightingale, P. D., G. Malin, C. S. Law, A. J. Watson, P. S. Liss, M. I. Liddicoat, J.
 1029 Boutin, and R. C. Upstill-Goddard (2000), In situ evaluation of air-sea gas exchange
 1030 parameterizations using novel conservative and volatile tracers., *Global*
 1031 *Biogeochemical Cycles*, 14, 373-387.

- 1032 Pandis, S. N., L. M. Russell, and J. H. Seinfeld (1994), The relationship between
 1033 DMS flux and CCN concentration in remote marine regions., *Journal of Geophysical*
 1034 *Research*, 99, 16,945-916,957.
- 1035 Saltzman, E. S., D. B. King, K. Holmen, and C. Leck (1993), Experimental
 1036 determination of the diffusion coefficient of dimethylsulfide in water., *Journal of*
 1037 *Geophysical Research*, 98, 16,481-416,486.
- 1038 Saxena, V. K., and S. Menon (1999), Sulfate-induced cooling in the southeastern
 1039 U.S.: An observational assessment., *Geophysical Research Letters*, 26, 2489-2492.
- 1040 Schwartz, S. E., and A. Slingo (1996), Enhanced shortwave cloud radiative forcing
 1041 due to anthropogenic aerosols., in *Clouds, Chemistry and Climate*, edited by P. J.
 1042 Crutzen and V. Ramanathan, pp. 191-235, Springer-Verlag.
- 1043 Simo, R. (2001), Production of atmospheric sulfur by oceanic plankton:
 1044 biogeochemical, ecological and evolutionary links., *Trends in Ecology and Evolution*,
 1045 16, 287-294.
- 1046 Simo, R., and J. Dachs (2002), Global ocean emission of dimethylsulfide predicted
 1047 from biogeophysical data., *Global Biogeochemical Cycles*, 16, 26-21:26-10.
- 1048 Simo, R., and C. Pedros-Alio (1999), Role of vertical mixing in controlling the
 1049 oceanic production of dimethyl sulphide, *Nature*, 402, 396-399.
- 1050 Sunda, W., D. J. Kieber, R. P. Kiene, and S. Huntsman (2002), An antioxidant
 1051 function for DMSP and DMS in marine algae., *Nature (London)*, 418, 317-320.
- 1052 Turner, S. M., M. Harvey, C. S. Law, P. D. Nightingale, and P. S. Liss (2004), Iron-
 1053 induced changes in oceanic sulfur biogeochemistry., *Geophysical Research Letters*,
 1054 31, doi:10.1029/2004GL020296.
- 1055 Turner, S. M., P. D. Nightingale, L. J. Spokes, M. I. Liddicoat, and P. S. Liss (1996),
 1056 Increased dimethylsulphide concentrations in sea water from in situ iron enrichment.,
 1057 *Nature*, 383, 513-517.
- 1058 Walsh, J., D. A. Dieterle, and J. Lenos (2001), A numerical analysis of carbon
 1059 dynamics of the Southern Ocean phytoplankton community: the roles of light and
 1060 grazing in effecting both sequestration of atmospheric CO₂ and food availability to
 1061 larval krill., *Deep-Sea Research I*, 48, 1-48.
- 1062 Wolfe, G. V., M. Steinke, and G. O. Kirst (1997), Grazing-activated chemical defence
 1063 in a unicellular marine alga., *Nature*, 387, 894-897.
- 1064 Zonneveld, C. (1998), Photoinhibition as affected by photoacclimation in
 1065 phytoplankton: a model approach., *Journal Of Theoretical Biology*, 193, 115-123.
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- 1067
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TABLES

Table 1: Parameters and values for the NPZ-DMS model.

PAR	PROCESS	UNITS	VALUES	
			G99	GA
k_1	Maximum rate of N uptake by P	d^{-1}	0.27	0.9918
k_2	Half-saturation constant for P uptake of N	mgNm^{-3}	12.6	19.1230
k_3	Z grazing rate (per individual) on P	$\text{m}^3\text{mgN}^{-1}\text{d}^{-1}$	0.02	0.0256
k_4	Proportion of N uptake excreted by Z	-	0.3	0.0287
k_5	Z specific mortality rate	d^{-1}	0.050	0.3720
m_1	Rate of release of $DMSP$ by P	d^{-1}	0.01	0.0076
m_2	Rate of $DMSP$ excretion rate by Z	d^{-1}	0.01	0.3050
m_3	$DMSP - DMS$ conversion rate	d^{-1}	0.5	0.9226
m_4	Rate of $DMSP$ consumption by bacteria	d^{-1}	0.53	0.1201
m_5	Rate of release of DMS by P	d^{-1}	0.0085	0.0045
m_6	Rate of DMS consumption by bacteria	d^{-1}	0.29	0.2664
m_7	Maximum DMS photo-oxidation rate	d^{-1}	1.3	0.5752
γ	Phytoplankton $S(DMSP):N$ ratio	mg S/mg N	0.3	0.3580
N_T	Total nutrient as dissolved inorganic nitrogen	mgNm^{-3}	50	3.0761

G99 represents Gabric et al [1999]; GA represents derived by GA for model

validation in this paper.

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1075 **Table 2: Additional parameters derived for the biofeedback model.**

PAR.	PROCESS	UNITS	VALUE
k_6	<i>P</i> -DMS flux parameter	mgS m mgN ⁻¹ d ⁻¹	0.0128
k_7	<i>Z</i> -DMS flux parameter	mgS m mgN ⁻¹ d ⁻¹	0.0035
k_8	<i>CCN</i> -DMS flux ratio	m ² d mgS ⁻¹	710
k_9	<i>CCN</i> background level	-	15
k_{10}	N_C - <i>CCN</i> ratio	-	183
k_{11}	N_C - <i>CCN</i> regression constant	-	334
k_{12}	$\Delta\alpha$ - N_C ratio	-	0.075
k_{13}	Phytoplankton <i>PI</i> curve parameter	-	0 - ∞
N_0	$\Delta\alpha$ - N_C reference droplet number (N_0)	-	-

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FIGURE LEGENDS

Figure 1. Schematic diagram of the *NPZ-DMS* feedback model. Dotted lines indicate the components of the original GMSK model that have been omitted for this analysis.

Figure 2: Photosynthesis-irradiance curve described by equation (30). Solid line has $b = 0.10$, dotted line is $b = 1$ and dashed line is $b = 10$.

Figure 3. Forcing data for depth-averaged *NPZ-DMS* model validations: (a) sea surface temperature (SST); (b) mixed layer depth (MLD), (c) photosynthetically active radiation (PAR) and (d) sea surface wind speed (WIN) for the Southern Ocean region 60-65°S, 125-140°E.

Figure 4. Best fits derived for depth-averaged *NPZ-DMS* model validations: (a) model P (line) and SeaWiFS chlorophyll data (dots) and (b) model DMS (line) and Kettle et al [1999] DMS data (dots). Data is for the same Southern Ocean region as Figure 3.

Figure 5. Normalised return time surface for the *NPZ-DMS* model without feedback showing the times the system takes to return to its equilibrium state after perturbation.

1097 Figure 6. Normalised return time surface for the *NPZ-DMS* model with feedback
1098 showing the times the system takes to return to its equilibrium state after perturbation.

1099

1100 Figure 7. Normalised return time difference surface for the *NPZ-DMS* model with and
1101 without biogenic feedback. This surface is calculated by subtracting the return time
1102 surface of the *NPZ-DMS* model with feedback from the return time surface of the
1103 *NPZ-DMS* model with no feedback. This surface has been smoothed for clarity of
1104 presentation.

1105

1106 Figure 8. Sensitivity of feedback effect to first order parameters identified by the New
1107 Morris Method. Light region of bar is sensitivity to each parameter alone; dark region
1108 is the influence of each parameter in association with all other parameters (i.e.
1109 indicates the existence of second and higher order interaction effects).

1110

1111 Figure 9. Sensitivity of feedback effect to second order parameter interactions
1112 identified by the New Morris Method. Light region of bar is sensitivity to each
1113 parameter pair alone; dark region is the influence of each parameter pair in association
1114 with all other parameters (i.e. indicates the existence of third and higher order
1115 interaction effects).

1116

1117 Figure 10. The effect of time lags on the feedback for the return time of the *NPZ-DMS*
1118 feedback model. The feedback effect is measured as the proportion by which sum of

1119 non-feedback model's return time surface exceeds the sum of the feedback model's
1120 return time surface. The thick solid line is the feedback model with the default
1121 parameter set, the dotted line is with the maximum phytoplankton growth rate (k_1)
1122 doubled, the dashed line is with the zooplankton grazing rate (k_3) doubled and the thin
1123 solid line is with the zooplankton mortality rate (k_5) doubled.

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